

ANTIBODIES FOR HEALTHY GUT FUNCTION

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TAKE HOME MESSAGE

The gastrointestinal tract is colonized by trillions of microorganisms termed the microbiome. These organisms have pivotal roles in the immunological, metabolic, nutritional, and physiological functions of the host and help keep the body in immune homeostasis, immune balance.

The gastrointestinal tract is at the major intersection of the immune system and microorganisms. Antibodies, immunoglobulins, are large proteins produced by the gut-associated lymphoid tissues (GALT) found in the lining of the intestines.

These antibodies help limit the growth of gut microorganisms, and pathogens that enter via the oral route. When the specificity or levels of immunoglobulins produced by the intestines is in insufficient quantities to protect the young, administration of commercially-available antibodies may be warranted.

MICROBIOME

The gastrointestinal tract (Bauer et al., 2006) and skin (Oh et al., 2014) of animals are colonized by trillions of dynamic ecosystems of microorganisms (Bauer et al., 2006). In totality it is estimated that these organisms express 10-fold more genes than does their host's genome. (Ley et al., 2006; Belkaid and Hand, 2014). As a community, the microorganisms in and on the body are termed the microbiome, and play key roles in the immunological, metabolic, nutritional, and physiological functions of the host (Bäckhed et al., 2005; Wu and Wu, 2012; Scott et al., 2013; Ji and Nielsen, 2015).

The regulatory functions of the microbiome are so vital that the microbiome is considered by some as a separate organ (O'Hara and Shanahan, 2006; Baquero and Nombela, 2012; Ji and Nielsen, 2015).

HOMEOSTASIS OF IMMUNE AND INTESTINAL FUNCTIONS

The intestine is constantly exposed to fluids, foodstuffs, microorganisms, and inorganic (Hooper et al., 2002) and organic materials. Among its many functions are: obtain nutrients, protect the host from infection, and modulate immunological homeostasis of the host (Hooper and Macpherson, 2010; Hooper et al., 2012; Wu and Wu, 2012; Brown et al., 2013; Levesque, 2014).

The lining of the intestine is made up of a single layer of epithelial cells which is highly permeable to permit fluids and nutrients to be transferred to the blood. The gastrointestinal tract has a complex system of immune cells immediately below the epithelium (Tomasello and Bedoui, 2013) that produces antibodies that inhibit the attachment of pathogens to the gut wall and help keep the intestines in microbial and immune homeostasis.

The importance of the gastrointestinal system for defense against orally-acquired infections is supported by the presence of high concentrations of immune cells in the intestines. About 70% of the immune system is represented within the gut-associated lymphoid tissues (GALT) (Jung et al., 2010; Vighi et al., 2010) with 80% of the body's immunoglobulin-producing plasma cells (Vighi et al., 2010) Peyer's patches, residing here as well.

As part of the digestive system's protective mechanisms, when the gut is exposed to antigenic pathogens, the plasma cells will produce immunoglobulins, antibodies, specific to the organisms that stimulated the response. Immunoglobulins, antibodies, are large Y-shaped proteins that bind to the antigens that initially triggered their production. Immunoglobulins attach to and "mark" the pathogen for destruction by immune factors and cells. Antibodies also inhibit the binding of microbes to the intestinal walls, decreasing the likelihood of infection.

There is constant interaction between the members of the gastrointestinal microbiome, immune networks (Shirkey, et al., 2006; Nicholson et al., 2012; Ji and Nielsen, 2015), and the intestinal epithelium cells (Shulzhenko, et al., 2011; de Vrese and Schrezenmeir, 2007; Goto and Ivanov, 2013). Exposure to gut microflora is essential for the full development of the immune structures in the intestines (Tomasello and Bedoui, 2013), and the microbes also have major influences on the structure and function of Peyer's patches (Barman et al., 1997) and other immune components of the gut.

Significant bi-directional influences between the microbiota and the immune system (Tomasello and Bedoui, 2013) play pivotal roles in defending the body from infection and mutating cells, and maintaining the host's immune and physiological homeostasis (Mason et al., 2008; Wu and Wu, 2012; Goto and Ivanov, 2013; Belkaid and Hand, 2014).

Imbalances in concentrations and composition (Belkaid and Hand, 2014) of microbiota affect immunity and local and systemic inflammation in areas distal from the intestines (Ichinohe, et al., 2011; Hand et al., 2012; Belkaid and Naik, 2013; Iida et al., 2013; Belkaid and Hand, 2014). As an example, broad-spectrum oral antibiotic treatment results in changes of the gut flora and influences immunological responsiveness to infection with influenza (Ichinohe et al., 2011).

IMMUNE PROTECTION IN THE NEWBORN

Born agammaglobulinemic, without significant amounts of protective antibodies (Hurley and Theil, 2011), newly born mammals, such as pigs and cows, are vulnerable to infection (Bauer et al., 2006) and other environmental challenges until their own immune systems and microbiome mature.

Initial protection from infection is through the first milk, colostrum, which is passively transferred from the mother (Bauer et al., 2006; Langer, 2009) for the first few days after birth. As the newborn's immune system matures and continues to nurse, the concentration of immunoglobulins and proteins provided in mother's milk decreases (Langer, 2009).

Ingestion of colostrum by the young significantly affects development of the gastrointestinal tract (Blum, 2006) and its microbiome (Cabrera-Rubio, et al., 2012). In pigs and cows, for the first 12-24 hours of life, whole antibodies move easily from the intestine into the circulatory system without being digested by enzymes. However within 24-36 hours after birth, “closure” of the digestive system occurs, and intestinal cells become selective in what passes through the gut wall (Staley and Bush, 1985; Blum, 2006; Hurley and Theil, 2011).

Until the newborn’s immune and microbiota has developed sufficiently for it to protect itself, the newly born must have adequate amounts of protective antibodies to meet challenges from invading pathogens. If the antigenic challenge is too overwhelming, or the antibody titers are not sufficient or the specificity of the antibodies not appropriate for the challenge, there may not be enough passively acquired antibody to adequately protect the newborn. Insufficient amounts of antibodies require that the young be cross-fostered onto another mother, or given supplemental antibodies (ThePigSite.com).

HETEROLOGOUS TRANSFER OF PASSIVE IMMUNITY

Typically, immunoglobulin-rich colostrum, hyperimmune milk, or hyperimmune eggs are used in humans and livestock to help the newborn defend itself from pathogenic challenges until its own immune and microbiota systems are mature.

A useful extension of our knowledge of passive transfer of protective immunoglobulins is the opportunity to use immunoglobulins from one host for prevention or treatment of disease in a secondary host.

Decades ago, clinicians orally transferred anti-rotavirus human immunoglobulin to three children with primary immunodeficiency syndromes that were suffering from chronic excretion of gastrointestinal rotavirus. The antibody survived passage through the gut and retained its ability to attach to rotavirus. Antigen-antibody complexes were found in the stool for 4-6d post administration of a single dose. For several days after treatment no viral antigen was detected in stools (Losonsky, et al., 1985).

Significant immune protection can also be transferred orally by using antibodies from one species for another (Hammarström et al., 1994; Kovacs-Nolan and Mine, 2004; Hurley and Theil, 2011). Immunoglobulins are fairly resistant to digestion and maintain their immunological activity (Losonsky, et al., 1985; Vega, et al., 2011). Even when immunoglobulins are enzymatically digested, the Fab2 and Fab fragments are still able to bind to the antigen and continue to have neutralizing properties. (Akita and Nakai, 1998; Carlander, 2002).

Hyperimmune products provide a high level of targeted protection to animals. Such proteins are typically produced by stimulating chickens or mammals multiple times with inactivated bacteria and/or viruses. In response to this massive antigenic exposure, the host produces both specific and non-specific immunoglobulins, along with high concentrations of other biological and immune factors that help maintain host immune homeostasis.

Hyperimmune proteins have been studied extensively in humans and other animals and shown to enhance functioning of joints, digestive and other organ systems (Heckert, et al., 1999; Mine and Kovacs-Nolan, 2002; Larsson and Carlander, 2003; Kelly GS, 2003/4; Schade R, et al., 2005; Hurley and Theil, 2011; Xu et al., 2011; Kramski et al., 2012).

EGG PROTEINS AS A SOURCE OF IMMUNOGLOBULINS AND OTHER IMMUNE FACTORS

The hen passively transfers immunity, nutrients, and growth factors to her chicks by means of the egg.

Eggs have been used since antiquity to enhance animal and human health. One of the oldest works on the medical treatment of donkeys, mules, and horses *P. Vegeti Renati Digestorum Artis Mulomedicinae libri* (Fischer, 2011) mentions egg as a remedy for diseases.

In human applications, the Roman army traveled extensively and had purpose-built hospitals with physicians trained and influenced by the Greeks. Chronicles suggest that raw eggs from the local areas were used to prevent and treat diseases such as dysentery (Lommatzsch, 1903) as the Roman army moved from territory to territory.

In response to antigenic stimulation of hens, either via environmental exposure to pathogens, or injection of microorganisms, antibodies are concentrated in the yolk, along with a wide-spectrum of bioactive elements which are found both in the white and in the yolk of the egg.

Hyperimmune egg is not only rich in immunoglobulins and nutrients, but it contains a multiplicity of pro- and –anti-inflammatory molecules, interferons, chemokines, anti-viral and anti-bacterial biological factors (Wu et al., 2010). Investigators have calculated that there is a 30x greater concentration of small immune factors in a single egg from a “hyperimmunized” hen as compared to a regular egg.

The discovery that immunoglobulin was detectable in egg yolks of vaccinated hens, as well as in the blood, pre-dates the use of an antibiotics (Klemperer , 1893). Although functionally equivalent to mammalian IgG (He et al., 2014), the type of immunoglobulin produced by birds was deemed different enough from mammalian antibodies and was thus designated as “IgY” (Leslie and Clem, 1969).

IgY not found in mammals, but is the dominant class of immunoglobulin found in avian egg yolk. It has multiple advantages over mammalian antibodies. For example, mammalian-derived antibodies can trigger inflammatory processes when they interact with rheumatoid factor or complement. IgY does not interact with either of these immunological factors (Gottstein and Hemmeler, 1985; Schade et al., 1991).

Another important advantage of IgY is that for microorganisms, its avidity, i.e., the binding capacity of IgY, is much tighter than the avidity of mammalian antibodies. Additionally, since fats and proteins in egg help protect IgY from enzymatic degradation, chicken immunoglobulins are less susceptible to digestion than bovine antibodies. Also a higher degree of protection is

obtained with egg antibodies as compared with the same amount of mammalian immunoglobulins (Ikemori, et. al. 1997).

Despite the fact that chicken egg is an impressive source of antibodies (Tini et al., 2002), egg-derived immunoglobulins are under-utilized for animal and human applications (Kovacs-Nolan J, Mine Y 2004; Carlander et al., 2002; Schade R, et al., 2005; Xu et al., 2011).

Oral administration of egg antibodies may be a natural means to reduce or eliminate the use of antibiotics and control infections from bacteria and viruses (Kollberg et. al. 2003; Larsson and Carlander, 2003; Kovacs-Nolan and Mine, 2004).

A partial list of pathogens against which IgY antibodies have been produced are: *Helicobacter pylori* (Shin et al., 2002; Wang et al., 2014; Yang et al., 2012), influenza (Yang et al., 2014), *Pseudomonas aeruginosa* infections in cystic fibrosis patients (Kollberg et al., 2003), hemorrhagic and bovine and human rotavirus viruses (Yolken et al., 1988; Kuroki et al., 1997; Sarker et al., 2001; Kovacs-Nolan and Mine, 2004; Li et al., 2014),), canine parvovirus-2 (Van Nguyen et al., 2006), infectious bursal disease (Yousif et al., 2006), *Toxoplasma* (Ferreira Júnior et al., 2012) and *Trypanosoma* (Sampaio et al., 2014).

Egg-derived IgY have also been studied as a less arduous means of producing antibodies against bovine and human diarrheas (Yokoyama et al., 1993; Erhard et al., 1993; Ikemori et al., 1997; Kweon et al., 2000; Vega et al., 2011; Diraviyam et al., 2014), *H. pylori*-induced gastritis (Shin et al., 2002), *Escherichia coli* (Akita and Nakai, 1998), and botulism neurotoxins (You et al., 2014), and snake (Aguilaret al., 2014), spider and scorpion venoms (Schade et al., 2005).

Orally-administered IgY is transferred and absorbed into the circulatory system of a piglet as efficiently as are the IgG antibodies from the sow's colostrum (Yokoyama et al., 1993). Immunoglobulins remain detectable in the neonatal circulation for 24-48h. Additionally, as seen above, specifically-induced egg yolk antibodies are protective against diarrhea in pigs (Yokoyama et al., 1993; Diraviyam et al., 2014) and other mammals.

As a side-benefit, administration of hyperimmune egg has been reported to contribute significantly in daily increases in weight gain (Heckert et al., 1999; Ikemori et al., 1997). Weight gains may be the result of the consumed immunoglobulins helping to control bioburdens in the lumen of the gut requiring less expenditure of energy to maintain gut homeostatic balance.

CONCLUSION

It is becoming increasingly evident that the gastrointestinal system is not merely an organ for digestion of feedstuffs, but has a myriad of other functions. Antibodies produced by the gut-associated lymphoid tissues, and immunoglobulins introduced exogenously, serve to protect the host from pathogens and partner with the gut microbiota and the immune system to achieve homeostatic balance.

WORKS CITED

- Aguilar I, Sánchez EE, Girón ME, Estrella A, Guerrero B, Rodriguez-Acosta FA. Coral snake antivenom produced in chickens (*Gallus domesticus*). *Rev Inst Med Trop Sao Paulo* 2014; 56: 61-66.
- Akita EM, Nakai S. Neutralization of enterotoxigenic *Escherichia coli* heat-labile toxin by chicken egg yolk immunoglobulin Y and its antigen-binding fragments. *Food Agric Immunol* 1998; 10: 161-172.
- Bäckhed F, Ley RE, Sonnenburg JL, Peterson DA, Gordon JI. Host-bacterial mutualism in the human intestine. *Science* 2005; 307: 1915-1920.
- Barman NN, Bianchi AT, Zwart RJ, Pabst R, Rothkötter HJ. Jejunal and ileal Peyer's patches in pigs differ in their postnatal development. *Anat Embryol (Berl)* 1997; 195: 41-50.
- Baquero F, Nombela C. The microbiome as a human organ. *Clin Microbiol Infect* 2012; S4: 2-4.
- Bauer E, Williams BA, Smidt H, Verstegen MW, Mosenthin R. Influence of the gastrointestinal microbiota on development of the immune system in young animals. *Curr Issues Intest Microbiol* 2006; 7: 35-51.
- Belkaid Y, Hand T. Role of the microbiota in immunity and inflammation. *Cell* 2014; 157: 121-141.
- Belkaid Y, Naik S. Compartmentalized and systemic control of tissue immunity by commensals. *Nat Immunol* 2013; 14: 646-653.
- Blum JW. Nutritional physiology of neonatal calves. *J Anim Physiol Anim Nutr* 2006; 90: 1–11.
- Brown EM, Sadarangani M, Finlay BB. The role of the immune system in governing host-microbe interactions in the intestine. *Nat Immunol*. 2013; 14: 660-667.
- Cabrera-Rubio R¹, Collado MC, Laitinen K, Salminen S, Isolauri E, Mira A. The human milk microbiome changes over lactation and is shaped by maternal weight and mode of delivery. *Am J Clin Nutr* 2012; 96: 544-551.
- Carlander D. Avian IgY Antibody: In vitro and in vivo. In: *Comprehensive Summaries of Uppsala Dissertations from the Faculty of Medicine 1119*. Acta Universitatis Upsaliensis Uppsala, Sweden. 2002; pp. 7-53.
- Diraviyam T, Zhao B, Wang Y, Schade R, Michael A, Zhang X. Effect of chicken egg yolk antibodies (IgY) against diarrhea in domesticated animals: a systematic review and meta-analysis. *PLoS One* 2014; 9: e97716.
- Erhard MH, Kellner J, Eichelberger J, Lösch U. New possibilities in oral immunoprophylaxis of newborn diarrhea in calves--a field study using specific egg antibodies. [In German] *Berl Munch Tierarztl Wochenschr* 1993; 106: 383-387.

Ferreira Júnior Á, Santiago FM, Silva MV, Ferreira FB, Macêdo Júnior AG, Mota CM, Faria MS, Silva Filho HH, Silva DA, Cunha-Júnior JP, Mineo JR, Mineo TW. Production, characterization and applications for *Toxoplasma gondii*-specific polyclonal chicken egg yolk immunoglobulins. *PLoS One* 2012; 7: e40391.

Fischer, K-D. "Mulomedicina Chironis." Brill's New Pauly Supplements I - Volume 2 : Dictionary of Greek and Latin Authors and Texts. Edited by: Manfred Landfester, in collaboration with Brigitte Egger. Brill Online, 2015. Reference. 05 August 2015
http://referenceworks.brillonline.com/entries/brill-s-new-pauly-supplements-i-2/mulomedicina-chironis-COM_0148> First appeared online: 2011.

Goto Y, Ivanov II. Intestinal epithelial cells as mediators of the commensal–host immune crosstalk. *Immunol Cell Biol* 2013; 91: 204–214.

Gottstein B, Hemmeler E. Egg yolk immunoglobulin Y as an alternative antibody in the serology of echinococcosis. *Z Parasitenkd* 1985; 71: 273-276.

Hammarström L, Gardulf A, Hammarström V, Janson A, Lindberg K, Smith CI. Systemic and topical immunoglobulin treatment in immunocompromised patients. *Immunol Rev* 1994; 139: 43-70.

He JX, Thirumalai D, Schade R, Zhang XY. Chronobiological studies of chicken IgY: monitoring of infradian, circadian and ultradian rhythms of IgY in blood and yolk of chickens. *Vet Immunol Immunopathol* 2014; 160: 266-272.

Heckert HP, Bardella I, Hofmann W, Oltmer S. Effect of antibody-containing egg powder on development of active immunity in calves. [In German] *Dtsch Tierarztl Wochenschr* 1999; 106: 10-14.

Hooper LV, Littman DR, Macpherson AJ. Interactions between the microbiota and the immune system. *Science* 2012; 336: 1268–1273.

Hooper LV, Macpherson AJ. Immune adaptations that maintain homeostasis with the intestinal microbiota. *Nat Rev Immunol* 2010; 10: 159–169.

Hooper LV, Midtvedt T, Gordon JI. How host-microbial interactions shape the nutrient environment of the mammalian intestine. *Annu Rev Nutr* 2002; 22: 283-307.

Hurley WK, Theil PK. Perspectives on immunoglobulins in colostrum and milk. *Nutrients* 2011; 3: 442–474.

Ichinohe T, Pang IK, Kumamoto Y, Peaper DR, Ho JH, Murray TS, et al. Microbiota regulates immune defense against respiratory tract *influenza A* virus infection. *Proc Natl Acad Sci USA* 2011; 108: 5354–5359.

Ikemori Y, Ohta M, Umeda K, Icatlo FC Jr, Kuroki M, Yokoyama H, Kodama Y. Passive protection of neonatal calves against bovine coronavirus-induced diarrhea by administration of egg yolk or colostrum antibody powder.

Vet Microbiol 1997; 58:105-111.

Iida N, Dzutsev A, Stewart CA, Smith L, Bouladoux N, Weingarten RA, Molina DA, Salcedo R, Back T, Cramer S, Dai RM, Kiu H, Cardone M, Naik S, Patri AK, Wang E, Marincola FM, Frank KM, Belkaid Y, Trinchieri G, Goldszmid RS. Commensal bacteria control cancer response to therapy by modulating the tumor microenvironment. *Science* 2013; 342: 967-970.

Ji B, Nielsen J. From next-generation sequencing to systematic modeling of the gut microbiome. *Front Genet* 2015; 6: Article 219: 1-9.

Jung C, Hugot, J-P, Barreau F. Review Article. Peyer's Patches: The Immune Sensors of the Intestine. *Int J Inflam* 2010; 2010: 1-12.

Kelly GS. Bovine colostrums: a review of clinical uses. *Altern Med Rev* 2003; 8:378-394. *Erratum in: Altern Med Rev* 2004; 9:69.

Klemperer F. Phylogeny of Immunoglobulin Structure and Function III. Immunoglobulins of the chicken. [In German] *Arch Exp Path Pharm* 1893; 31: 356-382.

Kollberg H, Carlander D, Olesen H, Wejåker PE, Johannesson M, Larsson A. Oral administration of specific yolk antibodies (IgY) may prevent *Pseudomonas aeruginosa* infections in patients with cystic fibrosis: a phase I feasibility study. *Pediatr Pulmonol* 2003; 35: 433-440.

Kovacs-Nolan J, Mine Y. Avian egg antibodies: basic and potential applications.

Avian Poult Biol Rev 2004; 15: 24-46.

Kramski M, Center RJ, Wheatley AK, Jacobson JC, Alexander MR, Rawlin G, Purcell DF. Hyperimmune bovine colostrum as a low-cost, large-scale source of antibodies with broad neutralizing activity for HIV-1 envelope with potential use in microbicides. *Antimicrob Agents Chemother* 2012; 56: 4310-9.

Kuroki M¹, Ohta M, Ikemori Y, Icatlo FC Jr, Kobayashi C, Yokoyama H, Kodama Y. Field evaluation of chicken egg yolk immunoglobulins specific for bovine rotavirus in neonatal calves. *Arch Virol* 1997; 142: 843-51.

Kweon CH, Kwon BJ, Woo SR, Kim JM, Woo GH, et al. Immunoprophylactic effect of chicken egg yolk immunoglobulin (IgY) against porcine epidemic diarrhea virus (PEDV) in piglets. *J Vet Med Sci* 2000; 62: 961-964.

Langer, P. Differences in the composition of colostrum and milk in eutherians reflect differences in immunoglobulin transfer. *J Mammal* 2009; 90: 332-339.

Larsson A, Carlander D. Oral immunotherapy with yolk antibodies to prevent infections in humans and animals. *Ups J Med Sci* 2003; 108: 129-140.

Leslie GA, Clem LW. Phylogeny of immunoglobulin structure and function. III. Immunoglobulins of the chicken. *J Exp Med* 1969; 130: 1337-1352.

Levesque CL. Immunology of the gut – taking the good with the bad. *Proc 75th MN Nutr Conf* 2014; pp. 9-16.

Ley RE, Peterson DA, Gordon JI. Ecological and evolutionary forces shaping microbial diversity in the human intestine. *Cell* 2006; 124: 837–848.

Li ZX, Hu WD, Li BC, Li TY, Zhou XY, Zhang Z. Egg yolk IgY against RHDV capsid protein VP60 promotes rabbit defense against RHDV infection. *Vet Immunol Immunopathol* 2014; 157: 97-104.

Lommatzsch, E. (Eds.) 1903. P. Vegeti Renati. Digestorum Artis, Mulomedicineae, Libri. Harvard College Library.

Losonsky GA, Johnson JP, Winkelstein JA, Yolken RH. Oral administration of human serum immunoglobulin in immunodeficient patients with viral gastroenteritis. A pharmacokinetic and functional analysis. *J Clin Invest* 1985; 76: 2362-2367.

Mason KL, Huffnagle GB, Noverr MC, Kao JY. Overview of gut immunology. *Adv Exp Med Biol* 2008; 635: 1-14.

Mine Y, Kovacs-Nolan J. Chicken egg yolk antibodies as therapeutics in enteric infectious disease: a review. *J Med Food* 2002; 5: 159-169.

Nicholson JK, Holmes E, Kinross J, Burcelin R, Gibson G, Jia W, Pettersson S. Host-gut microbiota metabolic interactions. *Science* 2012; 336: 1262–1267.

Nordqvist C. A History of Medicine. *Med News Today* 2012 August 9.

Oh J, Byrd AL, Deming C, Conlan S, NISC Comparative Sequencing Program, Kong HH,

Segre JA. Biogeography and individuality shape function in the human skin metagenome. *Nature* 2014; 514: 59-64.

O'Hara AM, Shanahan F. The gut flora as a forgotten organ. *EMBO Rep* 2006; 7: 688-693.

Sampaio LC, Baldissera MD, Grando TH, Gressler LT, Capeleto Dde M, de Sa MF, de Jesus FP, dos Santos AG Jr, Anciuti AN, Colonetti K, Stainki DR, Monteiro SG. Production, purification and therapeutic potential of egg yolk antibodies for treating *Trypanosoma evansi* infection. *Vet Parasitol* 2014; 29:96-103.

Sarker SA¹, Casswall TH, Juneja LR, Hoq E, Hossain I, Fuchs GJ, Hammarström L. Randomized, placebo-controlled, clinical trial of hyperimmunized chicken egg yolk immunoglobulin in children with rotavirus diarrhea. *J Pediatr Gastroenterol Nutr* 2001; 32: 19-25.

Schade R, Calzado EG, Sarmiento R, Chacana PA, Porankiewicz-Asplund J, Terzolo HR. Chicken egg yolk antibodies (IgY-technology): A review of progress in production and use in research and human and veterinary medicine. *Altern Lab Anim* 2005; 33: 129-54.

Schade R, Pfister C, Halatsch R, Henklein P. Polyclonal IgY antibodies from chicken egg yolk-an alternative to the production of mammalian IgG type antibodies in rabbits. *ATLA* 1991; 19: 403-419.

Scott KP, Gratz SW, Sheridan PO, Flint HJ, Duncan SH. The influence of diet on the gut microbiota. *Pharmacol Res* 2013; 69: 52-60.

Shin JH, Yang M, Nam SW, Kim JT, Myung NH, Bang WG, Roe IH. Use of egg yolk-derived immunoglobulin as an alternative to antibiotic treatment for control of *Helicobacter pylori* infection. *Clin Diagn Lab Immuno* 2002; 9: 1061-6.

Shirkey TW, Siggers RH, Goldade BG, Marshall JK, Drew MD, Laarveld B, Van Kessel AG. Effects of commensal bacteria on intestinal morphology and expression of proinflammatory cytokines in the gnotobiotic pig. *Exp Biol Med (Maywood)* 2006; 231: 1333-45.

Shulzhenko N¹, Morgun A, Hsiao W, Battle M, Yao M, Gavrilova O, Orandle M, Mayer L, Macpherson AJ, McCoy KD, Fraser-Liggett C, Matzinger P. Crosstalk between B lymphocytes, microbiota and the intestinal epithelium governs immunity versus metabolism in the gut. *Nat Med* 2011;17: 1585-1593.

Staley TE, Bush LJ. Receptor mechanisms of the neonatal intestine and their relationship to immunoglobulin absorption and disease. *J Dairy Sci* 1985; 68: 184–205.

ThePigSite.com. *Pig health. Acquired specific immunity*. Article 38. 5m Publishing, Benchmark House, 8 Smithy Wood Drive, Sheffield, S35 1QN, England.

Tini M, Jewell UR, Camenisch G, Chilov D, Gassmann M. Generation and application of chicken egg-yolk antibodies. *Comp Biochem Physiol A Mol Integr Physiol* 2002; 131: 569-74.

Tomasello E, Bedoui S. Intestinal innate immune cells in gut homeostasis and immunosurveillance. *Immunol Cell Biol* 2013; 91: 201–203.

Van Nguyen S, Umeda K, Yokoyama H, Tohya Y, Kodama Y. Passive protection of dogs against clinical disease due to *Canine parvovirus-2* by specific antibody from chicken egg yolk. *Can J Vet Res* 2006; 70: 62-64.

Vega C, Bok M, Chacana P, Saif L, Fernandez F, Parreño V. Egg yolk IgY: protection against rotavirus induced diarrhea and modulatory effect on the systemic and mucosal antibody responses in newborn calves. *Vet Immunol Immunopathol* 2011; 15: 156-69.

Vighi G, Marcucci F, Sensi L, Di Cara G, Frati F. Allergy and the gastrointestinal system. *Clin Exp Immunol* 2008; 153S 1:3-6.

de Vrese M, Schrezenmeir J. Effects of Probiotics and Prebiotics on Health Maintenance—Critical Evaluation of the Evidence- Preface to Supplement. *J Nutr* 2007; 137: 739S-740S.

Wang B, Yang J, Cao S, Wang H, Pan X, Zhu J, Zhou Y, Gao L, Li W, Li M. Preparation of specific anti-*Helicobacter pylori* yolk antibodies and their antibacterial effects. *Int J Clin Exp Pathol* 2014; 7: 6430-6437.

Wu HJ, Wu E. The role of gut microbiota in immune homeostasis and autoimmunity. *Gut Microbes* 2012; 3: 4-14.

Wu J, Majumder K, Gibbons K. 2010. Bioactive Proteins and Peptides from Egg Proteins. In: *Bioactive Proteins and Peptides as Functional Foods and Nutraceuticals*. pp. 247-264. Y. Mine, E. Li-Chan and B Jiang. (Eds.) Blackwell Publishing Ltd. and Institute of Food Technologists.

Xu Y, Li X, Jin L, Zhen Y, Lu Y, Li S, You J, Wang L. Application of chicken egg yolk immunoglobulins in the control of terrestrial and aquatic animal diseases: a review. *Biotechnol Adv* 2011; 29: 860-8.

Yang YH, Park D, Yang G, Lee SH, Bae DK, Kyung J, Kim D, Choi EK, Son JC, Hwang SY, Kim YB. Anti-*Helicobacter pylori* effects of IgY from egg yolk of immunized hens. *Lab Anim Res* 2012; 28: 55-60.

Yang YE, Wen J, Zhao S, Zhang K, Zhou Y. Prophylaxis and therapy of pandemic H1N1 virus infection using egg yolk antibody. *J Virol Methods* 2014; 206:19-26.

Yokoyama H, Peralta RC, Sendo S, Ikemori Y, Kodama Y. Detection of passage and absorption of chicken egg yolk immunoglobulins in the gastrointestinal tract of pigs by use of enzyme-linked immunosorbent assay and fluorescent antibody testing. *Am J Vet Res* 1993; 54: 867–872.

Yolken RH, Leister F, Wee SB, Miskuff R, Vonderfecht S. Antibodies to rotaviruses in chickens' eggs: a potential source of antiviral immunoglobulins suitable for human consumption. *Pediatrics* 1988; 81: 291-295.

You Z, Yang H, Xin W, Kang L, Gao S, Wang J, Zhang T, Wang J. Preparation of egg yolk antibodies against BoNT/B and their passive protection in mouse models. *Hum Vaccin Immunother* 2014; 10: 2321-7.

Yousif AA, Mohammad WA, Khodeir MH, Zeid AZ, el-Sanousi AA, Saber MS, Reda IM. Oral administration of hyperimmune IgY: an immunoeological approach to curbing acute infectious bursal disease virus infection. *Egypt J Immunol* 2006; 13: 85-94.